



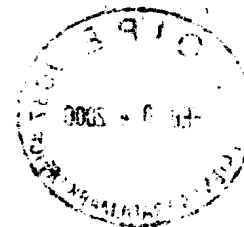
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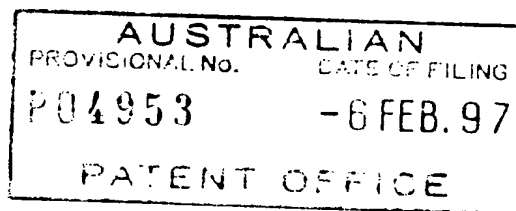
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I, LEANNE MYNOTT, TEAM LEADER EXAMINATION SUPPORT AND SALES hereby certify that annexed is a true copy of the Provisional specification in connection with Application No. PO 4953 for a patent by THE NATIONAL UNIVERSITY OF SINGAPORE filed on 06 February 1997.

WITNESS my hand this  
Twenty-ninth day of November 1999

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The National University of Singapore

**A U S T R A L I A**

**Patents Act 1990**

**PROVISIONAL SPECIFICATION**

for the invention entitled:

"Diagnosis of Parasites"

The invention is described in the following statement:



## DIAGNOSIS OF PARASITES

The present invention relates generally to novel genetic sequences isolated from a parasitic protozoa which infects humans and other animals and the uses of said sequences as  
5 diagnostic agents for the detection of said protozoa in a biological sample. In particular, the present invention provides genetic sequences of the extrachromosomal genetic element of the malaria agent *Plasmodium berghei* and synthetic oligonucleotide derivatives, homologues, analogues and fragments thereof. The genetic sequences of the present invention are particularly useful in the diagnosis, prophylactic treatment and therapeutic treatment of  
10 humans and other animals which are capable of being infected by or are actually infected by protozoa such as *Plasmodium ssp.*, for example *P. falciparum*, *P. vivax*, *P. malariae*, *P. ovale*, *P. cynomolgi*, *P. gonderi*, *P. (Hepatocytis) kochi*, *P. inui*, *P. knowlesi*, *P. reichenowi*, *P. rodhaini*, *P. schwetzi*, *P. cathemerium*, *P. elongatum*, *P. relictum*, *P. lophurae*, *P. gallinaceum*, *P. yoelii*, or *P. berghei*, amongst others. The invention provides further, a  
15 novel, reliable diagnostic assay for the detection of *Plasmodium ssp.* in humans and animals.

Bibliographic details of the publications referred to by author in this specification are collected at the end of the description. Sequence identity numbers (SEQ ID Nos.) for the  
20 nucleotide and amino acid sequences referred to in the specification are defined after the bibliography.

Throughout the specification, unless the context requires otherwise, the word "comprise", or variations such as "comprises" or "comprising", will be understood to imply  
25 the inclusion of a stated element or integer or group of elements or integers, but not the exclusion of any other element or integer or group of elements or integers.

More than fifty different species of *Plasmodium* can cause malaria in humans, monkeys, birds, fish, cattle and rodents. The development of diagnostic assays for the  
30 detection of *Plasmodium* in humans and animals is therefore highly desirable.

- 3 -

Human malaria, which is caused by *Plasmodium ssp.*, in particular *P. falciparum*, *P. vivax*, *P. malariae*, and *P. ovale*, remains one of the major health problems around the world.

*Plasmodium vivax* induces a moderate form of malaria, vivax malaria, characterized by periodic chills and fever, an enlarged spleen, anaemia, severe abdominal pain and headaches, and extreme lethargy. If left untreated, the disease tends to be self-limiting within a period of 10 to 30 days, but will recur periodically. Although the fatality rate of vivax malaria is low, the disease is highly debilitating and makes the patient more vulnerable to other diseases.

10

The incubation period ranges from 10 days to 4 weeks. Generally, paroxysms of chills and fever appear on the 14th day after the bite of an infected female anopheles mosquito. During this time the parasite has been multiplying in the liver cells of the patient. Paroxysms continue to recur every other day, as the parasite completes its 48-hour cycle of development, now in the blood. During the paroxysm, the patient first goes through a "cold stage" during which he has chilly sensations, his skin is blue, his teeth chatter and there is violent shaking. After an hour, the "hot stage" is ushered in, with a rise in temperature to as high as 107°F (41.7°C); the skin is hot and dry and the patient complains of severe headache. The fever lasts about 2 hours, and is followed by the "sweating stage", during which there is profuse perspiration, the temperature falls to normal, the headache disappears, and although weak and drowsy, the patient feels well.

20

*Plasmodium ovale* produces a disease very similar to vivax malaria.

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*Plasmodium malariae*, the causative agent of quartan malaria, has an incubation period of 18-40 days. The paroxysms occur every 72 hours, and are longer and somewhat more severe than those accompanying vivax malaria.

- 4 -

*Plasmodium falciparum*-induced malaria (falciparum malaria) presents oedema of the brain and lungs and blockage of the kidneys, in addition to the symptoms associated with vivax malaria. Unless treated promptly, the fatality rate of falciparum malaria is high, especially in juveniles.

5

Paroxysms associated with falciparum malaria occur irregularly after a 12-day incubation period. They are severe, and accompanied by high temperatures. The so-called cerebral algid, haemorrhagic and pernicious types of malaria represent forms of falciparum malaria with different localizations of the parasite. In the cerebral type, the onset is delirium  
10 and coma, and death may occur in several hours without return to consciousness. "Black-water fever" or haemorrhagic malaria is a type in which haemolysis or dissolution of the red cells occurs, and dark urine due to the presence of haemoglobin is an outstanding feature. In the algid form, there are vomiting, diarrhea, and subnormal temperature.

15 The life cycle of the parasite and its course in the human body proceeds in the following way. The saliva of the mosquito contains the *Plasmodium* at the lance-shaped sporozoite stage of its life cycle. Upon inoculation of the host by biting, the sporozoites quickly migrate to the liver where they divide and develop into multi nucleated schizonts. Within 6 to 12 days, the schizonts disrupt and release into the blood the form known as  
20 *merozoites*. Each liver cell infected by one sporozoite releases into the blood stream from 10,000 to 30,000 merozoites. These later invade the host's erythrocytes where they grow and form more schizonts which, in turn, again divide, releasing more merozoites into the blood stream to repeat the cycle. The principal symptoms of malaria are associated with the rupture of the schizonts, the periodic lysis of the blood cells with release of merozoites and toxic  
25 wastes which cause the regular fevers and chills of malaria.

Neither vector control measures nor immuno or chemoprophylaxis have proven effective in eradicating the disease. Thus, more than ever, chemotherapy appears to be crucial in dealing with both the prevention and treatment of malaria. However, presently used drugs  
30 are constantly losing their efficacy due to the development of drug resistance by the parasite.

- 5 -

For example, drug resistance of *Plasmodium falciparum* to chloroquine has occurred in Bangladesh, Brazil, Burma, Colombia, Ecuador, Guyana (French), Guyana, India, Indonesia, Kampuchea, Malaysia, Nepal, Pakistan, Panama, Philippines, Surinam, Thailand, Venezuela, and Vietnam, amongst others. Therefore, the design of novel drugs is urgent.

5

Targets for drug design are generally nuclear-encoded gene products. However, inter-specific and developmental variation in nuclear gene expression has reduced the general efficacy of drugs which target such nuclear-encoded gene products.

10       Diagnosis of malaria is generally made by microscopic examination of blood films taken during episodes of fever, when the parasites may be seen. In general, the *Plasmodium* parasite is detected microscopically by examining finger prick blood samples for the presence of the morphologically distinct parasite using Giemsa stain solution (Shute *et al.*, 1980). This needs to be done by an experienced microscopist since *Plasmodium falciparum* and  
15 *Plasmodium vivax* are morphologically similar, albeit not identical. In view of the distinct epidemiologies of *P. falciparum* compared to *P. vivax*, it is important that diagnosis of infection by these species have a low error rate. Any incorrect diagnosis of falciparum malaria, for example, may be fatal for the patient. The microscopic technique is limited in so far as the method is slow and specialised personnel is required to perform the technique.

20

A variation of the standard microscopic assay, the quantitative buffy coat (QBC) technique is based upon the ability of parasite nucleoproteins to absorb acridine orange and fluoresce (Wardlaw *et al.*, 1983). The fluorescent nucleoproteins are readily visible against a background of non-fluorescent red blood cells. Although the method is more sensitive than  
25 the standard microscopic assay, it suffers from the disadvantages associated with the standard microscopic assay. Furthermore, the requirement of costly fluorescence microscopes and centrifuges to perform the QBC assay, renders the method unrealistic in resource-limited settings which often lack even electricity.

30       Immunological tests, for example the ParaSight™ F test (Becton Dickinson) and the



- 6 -

similar ICT Malaria P.f. test (ICT Diagnostics) detect the *Plasmodium falciparum* histidine-rich protein FRP2 in blood samples derived from patients. A major drawback associated with such methods is that they require *Plasmodium falciparum* gene expression to occur before the organism can be detected. Furthermore, as considerable variation in gene expression can occur between *Plasmodium ssp.*, these tests tend to be species-specific. For example, the ParaSight™ F test (Becton Dickinson) and ICT Malaria P.f. test (ICT Diagnostics) are specific for *Plasmodium falciparum* only and incapable of detecting other species. Furthermore, these tests, in particular the ParaSight™ F test (Becton Dickinson), are subject to a high proportion of false-negative detections, such that a higher than acceptable frequency of patients infected with a *Plasmodium ssp.* go undetected.

Immunological techniques such as the enzyme-linked immunosorbent assay (ELISA) or the radio immunoassay (RIA) which detect genus- and species-specific parasite antigens also exist. However, such methods are constrained by immunological cross-reaction between parasite and host antigens on the one hand and between parasite antigens and antigens derived from other microorganisms on the other hand. As a consequence, the susceptibility of immunological methods to false positive detection of *Plasmodium* is high. As already mentioned above, species-specific detection methods lead to a large number of false-negative detections.

20

Furthermore, as different *Plasmodium* antigens are expressed at different developmental stages, immunological techniques may only detect the parasite at certain stages of development. Such antigenic diversity displayed by *Plasmodium* is a major obstacle to the application of immunological techniques. In addition, radioisotope-based assays such as the RIA are impractical for field use. Immunological methods cannot distinguish between past and present infections.

State-of-the art diagnostic assays, which rely on the detection of *Plasmodium* genomic DNA in a sample, are species-specific and not capable of general application for any *Plasmodium ssp.*, in part because there is considerable variation in genomic DNA between

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*Plasmodium* species, such variation precluding the simultaneous detection of several *Plasmodium ssp.* in a single biological sample or alternatively, the use of a single DNA-based assay for the detection of any *Plasmodium ssp.* in a biological sample derived from a human or animal subject suspected of carrying the parasite.

5

As a consequence of the foregoing, there is a high demand for a reliable and simple technology for the diagnosis of *Plasmodium* in human and animal tissues.

*Plasmodium ssp.* possess additional genomes with potentially crucial functions  
10 (Wilson *et al.*, 1991). Until the present invention, very little was known about this extrachromosomal material. Furthermore, the function of the extrachromosomal plasmid-like element in the protozoans remains to be determined. To date, there is no clear evidence for DNA replication or functionally active gene products from the plasmid-like element.

15 In work leading up to the present invention, the inventors have discovered that the molecular composition, physical arrangement and nucleotide sequence of the extrachromosomal plasmid-like element is highly conserved in different *Plasmodium ssp.*

The inventors have utilised the high degree of homology between different  
20 *Plasmodium ssp.* in the design of reliable, genera-specific or species-specific diagnostic assays for the detection of *Plasmodium*. The diagnostic assays described herein provide a significant advantage over currently employed assays based upon the detection of *Plasmodium* genomic DNA.

25 Furthermore, the inventors have discovered that the assays described herein provide the added advantage of excluding the high frequency of false negative detection of *Plasmodium* in a biological sample to a greater degree than known diagnostics.

The inventors further contemplate the use of polypeptides encoded by the  
30 extrachromosomal plasmid-like element, and their homologues, analogues and derivatives,

- 8 -

as targets for drug design and in the development of anti-malarial vaccines.

Accordingly, one aspect of the invention provides a diagnostic assay for the detection of *Plasmodium* in a biological sample derived from a human or animal subject, said assay  
5 comprising the detection of a *Plasmodium* extrachromosomal genetic element or a homologue, analogue or derivative thereof in said sample.

In an alternative embodiment, the invention provides a diagnostic assay for the detection of *Plasmodium* in a biological sample derived from a human or animal subject, said  
10 assay comprising the steps of hybridising a *Plasmodium ssp.* extrachromosomal genetic element probe or a homologue, analogue or derivative thereof to said sample and then detecting said hybridisation using a detection means.

According to this aspect, the *Plasmodium* detected using the invention may be any  
15 species of *Plasmodium* which carries an extrachromosomal genetic element.

In a preferred embodiment, the *Plasmodium* being detected is selected from the list comprising *P. falciparum*, *P. vivax*, *P. malariae*, *P. ovale*, *P. cynomolgi*, *P. gonderi*, *P. (Hepatocytis) kochi*, *P. inui*, *P. knowlesi*, *P. reichenowi*, *P. rodhaini*, *P. schwetzi*, *P.*  
20 *cathemerium*, *P. elongatum*, *P. relictum*, *P. lophurae*, *P. gallinaceum*, *P. yoelii*, or *P. berghei*, amongst others.

In a more particularly preferred embodiment however, the present invention is useful for the detection of a *Plasmodium* in biological samples derived from humans and the  
25 *Plasmodium* in such cases is selected from the list comprising *P. falciparum*, *P. vivax*, *P. malariae* and *P. ovale*, amongst others.

The term "biological sample" as used herein shall be taken to refer to any organ, tissue, cell, exudate, nucleic acid, protein, nucleoprotein or other material which is derived  
30 from a living or once-living organism. Accordingly, a biological sample may be prepared in

a suitable solution, for example an extraction buffer or suspension buffer. The present invention extends to the diagnosis of biological solutions thus prepared, the only requirement being that said solution at least comprises a biological sample as described herein.

5       The biological sample to be tested according to the invention, is derived from a human or animal species, in particular a human or animal which is capable of being infected by a *Plasmodium*. A particular advantage of the present invention is that it may be readily adapted to facilitate the analysis of any biological sample derived from a human or other animal. Those skilled in the relevant art will know how to modify the assay of the invention for the  
10 purposes of adapting said assay to the analysis of different biological tissues, where relevant or indicated, without any undue experimentation.

In a particularly preferred embodiment, the biological sample may be derived from the blood tissue of a human or animal subject, or cells, nucleic acid molecules and exudates  
15 derived therefrom, for example buffy coat, plasma, DNA or RNA, amongst others.

The term "extrachromosomal genetic element" shall be taken to refer to any nucleic acid molecule, in particular DNA or RNA, which comprises a part of the complete genetic material of a *Plasmodium ssp.* but which does not comprise a part of a *Plasmodium ssp.*  
20 chromosome or a direct gene product thereof. An extrachromosomal genetic element of a *Plasmodium ssp.* may or may not replicate independently of the *Plasmodium* genome, such that the copy number of said genetic element may vary between *Plasmodium* cells.

Accordingly, a *Plasmodium* extrachromosomal genetic element as hereinbefore defined  
25 may be a linear or circular DNA molecule. In this regard, a linear DNA molecule may resemble, at the nucleotide sequence level at least, mitochondrial DNA (Suplick *et al*, 1988), while a circular DNA molecule in a *Plasmodium* may be a plasmid or plasmid-like molecule which may resemble a vestigial plastid genome (Gardner *et al*, 1991; Howe *et al*, 1992).

30       In a preferred embodiment of the invention, the *Plasmodium* extrachromosomal

- 10 -

genetic element is a plasmid or plasmid-like DNA molecule comprising approximately 30-35kb of nucleotides in length.

Furthermore, the diagnostic assay of the present invention is useful for the detection  
5 of a *Plasmodium* extrachromosomal genetic element or a *Plasmodium*-derived extrachromosomal genetic element, regardless of whether or not said genetic element expresses or is capable of expressing a polypeptide product.

The term "*Plasmodium*-derived" as used herein shall be taken to refer to an integer  
10 which, although it originates from a *Plasmodium ssp.* is not necessarily present in its natural state. For example, an extrachromosomal genetic element may be derived from a *Plasmodium ssp.* if it has been purified or partially purified and/or modified by digestion with restriction endonucleases or other DNA-modifying enzymes, to produce an analogue or derivative molecule.

15

The *Plasmodium* extrachromosomal genetic element probe may be a plasmid or plasmid-like molecule derived from a *Plasmodium ssp.* which is capable of infecting a human or animal subject.

20       Wherein the probe is a plasmid or plasmid-like extrachromosomal genetic element or a homologue, analogue or derivative thereof, it is preferred that it is derived from a species of *Plasmodium* other than *P. falciparum*.

In a particularly preferred embodiment, the plasmid or plasmid-like extrachromosomal  
25 genetic element probe or a homologue, analogue or derivative thereof, is derived from *Plasmodium berghei*.

However, in a more particularly preferred embodiment of the invention, the plasmid or plasmid-like extrachromosomal genetic element probe is at least 95% identical to one or  
30 more of the sequences set forth in SEQ ID NOs:1-10 or its complementary nucleotide

- 11 -

sequence, or a homologue, analogue or derivative thereof.

Alternatively, the plasmid or plasmid-like extrachromosomal genetic element probe is capable of hybridising under high stringency conditions to one or more of the sequences set forth in SEQ ID NOs:1-10 or a complementary nucleotide sequence or a homologue, analogue or derivative thereof.

In a further alternative embodiment, the *Plasmodium ssp.* extrachromosomal genetic element probe preferably comprises a sequence of nucleotides of at least 15 nucleotides, more preferably at least 25 nucleotides, even more preferably at least 50 nucleotides and even more preferably at least 100 nucleotides or 500 nucleotides derived from the sequence set forth in SEQ ID NOs:1-4 or its complement or a homologue, analogue or derivative thereof.

In a most particularly preferred embodiment, the plasmid or plasmid-like extrachromosomal genetic element probe comprises a nucleotide sequence set forth in any one or more of SEQ ID NOs:1-10 or a complementary nucleotide sequence, or a homologue, analogue or derivative thereof.

For the purposes of nomenclature, the nucleotide sequences set forth in SEQ ID NOs:1-4 correspond to one strand of the PSI-PL470, PLH-PPH, PRB and PWQ open reading frames, respectively, of the 30.7kb *plasmodium berghei* plasmid-like extrachromosomal genetic element. The inventors have shown that the extrachromosomal genetic element is transcriptionally-active, using reverse transcription polymerase chain reaction (RT-PCR), and encodes organelle-like rRNAs, tRNAs, ribosomal proteins and RNA polymerase subunits, amongst others.

The nucleotide sequences set forth in SEQ ID Nos:5-10 correspond to synthetic oligonucleotide sequences derived from the *Plasmodium* extrachromosomal genetic element of *P. berghei*.

- 12 -

For the present purpose, "homologues" of a nucleotide sequence shall be taken to refer to an isolated nucleic acid molecule which is substantially the same as the nucleic acid molecule of the present invention or its complementary nucleotide sequence, notwithstanding the occurrence within said sequence, of one or more nucleotide substitutions, insertions, 5 deletions, or rearrangements.

"Analogues" of a nucleotide sequence set forth herein shall be taken to refer to an isolated nucleic acid molecule which is substantially the same as a nucleic acid molecule of the present invention or its complementary nucleotide sequence, notwithstanding the 10 occurrence of any non-nucleotide constituents not normally present in said isolated nucleic acid molecule, for example carbohydrates, radiochemicals including radio nucleotides, reporter molecules such as, but not limited to biotin DIG, alkaline phosphatase or horseradish peroxidase, amongst others.

15 "Derivatives" of a nucleotide sequence set forth herein shall be taken to refer to any isolated nucleic acid molecule which contains significant sequence similarity to said sequence or a part thereof. Generally, the nucleotide sequence of the present invention may be subjected to mutagenesis to produce single or multiple nucleotide substitutions, deletions and/or insertions. Nucleotide insertional derivatives of the nucleotide sequence of the present 20 invention include 5' and 3' terminal fusions as well as intra-sequence insertions of single or multiple nucleotides or nucleotide analogues. Insertional nucleotide sequence variants are those in which one or more nucleotides or nucleotide analogues are introduced into a predetermined site in the nucleotide sequence of said sequence, although random insertion is also possible with suitable screening of the resulting product being performed. Deletional 25 variants are characterised by the removal of one or more nucleotides from the nucleotide sequence. Substitutional nucleotide variants are those in which at least one nucleotide in the sequence has been removed and a different nucleotide or nucleotide analogue inserted in its place.

30 The present invention encompasses all such homologues, analogues or derivatives of

a *Plasmodium* extrachromosomal genetic element, subject to the proviso that said homologues, analogues or derivatives are useful in the performance of at least one assay format as described herein.

5       The inventors have discovered that the extrachromosomal genetic element of *Plasmodium* is particularly useful as a marker of *Plasmodium* infection in a human or animal subject, because the detection of said element is not subject to the disadvantages of other assay methods, in particular the prevalence of false negative detection. As a consequence, fewer numbers of *Plasmodium*-infected hosts escape detection, by screening such hosts for  
10 the presence of the extrachromosomal genetic element according to the embodiments described herein (1% or less false negative detection compared to 3% or more for other methods), than by screening for the presence of other *Plasmodium*-expressed genes or by screening for the expression products of said genes.

15       The present invention clearly contemplates diagnostic assays which are capable of both genera-specific or species-specific detection. Accordingly, in one embodiment, the *Plasmodium ssp.* extrachromosomal genetic element probe or homologue, analogue or derivative thereof comprises DNA capable of being used to detect multiple *Plasmodium ssp.* In an alternative embodiment, the *Plasmodium ssp.* extrachromosomal genetic element probe  
20 or homologue, analogue or derivative thereof comprises DNA capable of being used to detect a particular *Plasmodium ssp.*

The inventors have discovered further that the coding region of a *Plasmodium* extrachromosomal genetic element is highly-conserved in different *Plasmodium ssp.*, while  
25 there is much more variation at the nucleotide level in the non-coding regions. Whilst not being bound by any theory or mode of action, the more highly conserved sequences in the extrachromosomal genetic element derived from a particular species of *Plasmodium* are particularly useful as genera-specific probes for the detection of any *Plasmodium*, while the less-conserved sequences of said element may be useful as species-specific probes for the  
30 detection of a sub-group of *Plasmodium*, for example a sub-group which infects humans or



primates as opposed to other animals, or which induces a specific form of malaria in humans.

Furthermore, the diagnostic assay of the present invention may also be adapted to a genera-specific or species-specific assay by varying the stringency of the hybridisation step.

5 Accordingly, a low or lower stringency hybridisation may be used to detect several different species of *Plasmodium* in one or more biological samples being assayed, while a high or higher stringency of hybridisation is used to detect the presence of a specific species of *Plasmodium*.

10 For the purposes of defining the level of stringency, a low stringency is defined herein as being a hybridisation and/or a wash carried out in 6xSSC buffer, 0.1 % (w/v) SDS at 28°C. A moderate stringency is defined herein as being a hybridisation and/or wash carried out in 2xSSC buffer, 0.1 % (w/v) SDS at a temperature in the range 45°C to 65°C. A high stringency is defined herein as being a hybridisation and/or wash carried out in 0.1xSSC  
15 buffer, 0.1 % (w/v) SDS at a temperature of at least 65°C. Those skilled in the art will be aware of equivalent reaction conditions to those described herein for defining the hybridisation stringency.

Generally, the stringency is increased by reducing the concentration of SSC buffer,  
20 and/or increasing the concentration of SDS and/or increasing the temperature of the hybridisation and/or wash. Those skilled in the art will be aware that the conditions for hybridisation and/or wash may vary depending upon the nature of the hybridisation membrane or the type of hybridisation probe used. Conditions for hybridisations and washes are well understood by one normally skilled in the art. For the purposes of clarification of the  
25 parameters affecting hybridisation between nucleic acid molecules, reference is found in pages 2.10.8 to 2.10.16. of Ausubel *et al.* (1987), which is herein incorporated by reference.

The detection means according to this aspect of the invention may be any nucleic acid-based detection means, for example nucleic acid hybridisation techniques or polymerase chain  
30 reaction (PCR). The invention further encompasses the use of different assay formats of said

nucleic acid-based detection means, including restriction fragment length polymorphism (RFLP), amplified fragment length polymorphism (AFLP), single-strand chain polymorphism (SSCP), amplification and mismatch detection (AMD), interspersed repetitive sequence polymerase chain reaction (IRS-PCR), inverse polymerase chain reaction (iPCR) and reverse  
5 transcription polymerase chain reaction (RT-PCR), amongst others.

Wherein the detection means is a nucleic acid hybridisation technique, the *Plasmodium* extrachromosomal genetic element probe may be labeled with a reporter molecule capable of producing an identifiable signal (e.g. a radioisotope such as  $^{32}\text{P}$  or  $^{35}\text{S}$  or a biotinylated  
10 molecule). According to this embodiment, those skilled in the art will be aware that the detection of said reporter molecule provides for identification of the *Plasmodium* extrachromosomal genetic element probe and that, following the hybridisation reaction, the detection of the corresponding *Plasmodium ssp.* extrachromosomal genetic element in the biological sample is facilitated.

15

Wherein the detection means is an RFLP, nucleic acid derived from the biological sample, in particular DNA, is digested with one or more restriction endonuclease enzymes and the digested DNA is subjected to electrophoresis, transferred to a solid support such as, for example, a nylon or nitrocellulose membrane, and hybridised to the *Plasmodium*  
20 extrachromosomal genetic element probe as hereinbefore defined, optionally labeled with a reporter molecule. According to this embodiment, a specific pattern of DNA fragments is hybridised to the *Plasmodium* extrachromosomal genetic element probe, said pattern optionally specific for a particular *Plasmodium ssp.*, to enable the user to distinguish between different species of the parasite.

25

Wherein the detection means is a polymerase chain reaction or a variant of same, one or more nucleic acid primer molecules of at least 15 contiguous nucleotides in length derivable from the *Plasmodium* extrachromosomal genetic element probe as hereinbefore defined, or its complementary nucleotide sequence or a homologue, analogue or derivative  
30 thereof, is hybridised to the biological sample comprising nucleic acid or alternatively, to

- 16 -

nucleic acid derived from said sample and nucleic acid copies of the *Plasmodium* extrachromosomal genetic element present in said sample or a part or fragment thereof are enzymically-amplified.

5        Those skilled in the art will be aware that there must be a sufficiently high percentage nucleotide sequence identity between the *Plasmodium* extrachromosomal genetic element probe(s) and the sequences in the template molecule to which it(they) hybridise. As stated previously, the hybridisation conditions may be varied to promote hybridisation.

10        Preferably, the *Plasmodium* extrachromosomal genetic element probe is at least 95 % identical to the complement of the nucleotide sequence in the template molecule to which it hybridises. More preferably, the *Plasmodium* extrachromosomal genetic element probe is substantially the same as the complement of the nucleotide sequence in the template molecule to which it hybridises.

15

Preferably, the *Plasmodium* extrachromosomal genetic element probe is contained in an aqueous mixture of other nucleic acid primer molecules. More preferably, the nucleic acid primer molecule is in a substantially pure form.

20        According to this embodiment of the invention, the *Plasmodium* extrachromosomal genetic element probe(s) may comprise inosine, adenine, guanine, thymidine, cytidine or uracil residues or functional analogues or derivatives thereof which are capable of being incorporated into a polynucleotide molecule, provided that the resulting probe is capable of hybridising under at least low stringency conditions to a *Plasmodium* extrachromosomal  
25 genetic element.

In a particularly preferred embodiment, the *Plasmodium* extrachromosomal genetic element probe comprises the sequence of nucleotides set forth in any one or more of SEQ ID Nos: 5-10 or a complementary strand or a homologue, analogue or derivative thereof.

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- 17 -

In a more particularly preferred embodiment, the *Plasmodium* extrachromosomal genetic element probes are hybridised to a *Plasmodium* extrachromosomal genetic element contained in the biological sample being analysed, as probe pairs, in the combinations comprising SEQ ID Nos:5-6 or SEQ ID Nos:7-8 or SEQ ID NOs:9-10 or complementary  
5 strands, homologues, analogues or derivatives thereof.

In a most particularly preferred embodiment, the *Plasmodium* extrachromosomal genetic element probes are hybridised to a *Plasmodium* extrachromosomal genetic element contained in the biological sample being analysed, as the L/L probe pair comprising SEQ ID  
10 Nos:5-6 or complementary strands, homologues, analogues or derivatives thereof.

The *Plasmodium* extrachromosomal genetic element present in the biological sample, or a part or fragment thereof which is enzymically-amplified, is defined herein as a "template molecule". The template molecule may be a genetic sequence which is at least 40% identical  
15 at the nucleotide sequence level to SEQ ID NOs:1-4 or to its complementary nucleotide sequence, the only requirement being that it comprises a *Plasmodium* extrachromosomal genetic element as hereinbefore defined.

Those skilled in the art will also be aware that, in one format, the polymerase chain  
20 reaction provides for the hybridisation of non-complementary *Plasmodium* extrachromosomal genetic element probes to different strands of the template molecule, such that the hybridised probes are positioned to facilitate the 5'→ 3' synthesis of nucleic acid in the intervening region, under the control of a thermostable DNA polymerase enzyme. As a consequence, the polymerase chain reaction provides an advantage over other detection means in so far as the  
25 nucleotide sequence in the region between the hybridised *Plasmodium* extrachromosomal genetic element probes may be unknown and unrelated to any known nucleotide sequence.

In a particularly preferred embodiment, the nucleic acid template molecule comprises, in addition to other nucleotide sequences, a sequence of nucleotides derived from or  
30 contained within any one or more of the sequences set forth in SEQ ID Nos:1-10 or a

- 18 -

complementary sequence or a homologue, analogue or derivative thereof.

In an alternative embodiment, wherein the detection means is AFLP, the *Plasmodium* extrachromosomal genetic element probes are selected such that, when nucleic acid derived  
5 from the biological sample, in particular DNA, is amplified, different length amplification products are produced from different *Plasmodium ssp.* The amplification products may be subjected to electrophoresis, transferred to a solid support such as, for example, a nylon or nitrocellulose membrane, and hybridised to the *Plasmodium* extrachromosomal genetic element probe as hereinbefore defined, optionally labeled with a reporter molecule. According  
10 to this embodiment, a specific pattern of amplified DNA fragments is hybridised to the *Plasmodium* extrachromosomal genetic element probe, said pattern optionally specific for a particular *Plasmodium ssp.*, to enable the user to distinguish between different species of the parasite in much the same way as for RFLP analysis.

15 The technique of AMD facilitates, not only the detection of a *Plasmodium* extrachromosomal genetic element in a biological sample, but also the determination of nucleotide sequence variants which differ from the *Plasmodium* extrachromosomal genetic element probe used in the assay format.

20 Wherein the detection means is AMD, the *Plasmodium* extrachromosomal genetic element probe is end-labeled with a suitable reporter molecule and mixed with an excess of the amplified template molecule. The mixtures are subsequently denatured and allowed to renature to form nucleic acid "probe:template hybrid molecules" or "hybrids", such that any nucleotide sequence variation between the probe and the template molecule to which it is  
25 hybridised will disrupt base-pairing in the hybrids. These regions of mismatch are sensitive to specific chemical modification using hydroxylamine (mismatched cytosine residues) or osmium tetroxide (mismatched thymidine residues), allowing subsequent cleavage of the modified site using piperidine. The cleaved nucleic acid may be analysed using denaturing polyacrylamide gel electrophoresis followed by standard nucleic acid hybridisation as  
30 described *supra* to detect the *Plasmodium* extrachromosomal genetic element nucleotide

sequences.

Those skilled in the art will be aware of the means of end-labeling a genetic probe according to the performance of the invention described in this embodiment.

5

According to this embodiment, the use of a single end-labeled probe allows unequivocal localisation of the sequence variation. The distance between the point(s) of sequence variation and the end-label is represented by the size of the cleavage product.

10 In an alternative embodiment of AMD, the probe is labeled at both ends with a reporter molecule, to facilitate the simultaneous analysis of both DNA strands.

Wherein the detection means is IRS-PCR, the *Plasmodium* extrachromosomal genetic element probes are selected such that they each include one highly-repetitive restriction enzyme cleavage site, for example *AluI*, which is ubiquitous in many genomes. According to this embodiment, the appropriate restriction enzyme cleavage site is selected such that it is ubiquitous in *Plasmodium* extrachromosomal genetic element nucleotide sequences. The amplified template DNA is electrophoresed under conditions which facilitate high resolution and optionally probed with a labeled *Plasmodium* extrachromosomal genetic element probe.

20

Optionally, the amplified template DNA may be end-filled using Klenow fragment of DNA polymerase I or other suitable means, prior to the electrophoresis step.

According to this embodiment, different combinations of probes produce different patterns of amplified template nucleic acid.

25 Furthermore, with any probe combination used, each *Plasmodium ssp.* will produce a distinctive pattern of amplified template nucleic acid. As a consequence, the detection means is suitable for distinguishing between different *Plasmodium ssp.*, in addition to being useful for the detection of the *Plasmodium* extrachromosomal genetic element *per se* in a

30

biological sample.

Wherein the detection means is RT-PCR, the nucleic acid sample comprises an RNA molecule which is a transcription product of the *Plasmodium* extrachromosomal genetic  
5 element DNA or a homologue, analogue or derivative thereof. As a consequence, this assay format is particularly useful when it is desirable to determine expression of one or more *Plasmodium* extrachromosomal genetic element genes.

According to this embodiment, the RNA sample is reverse-transcribed to produce the  
10 complementary single-stranded DNA which is subsequently amplified using standard procedures.

The present invention clearly extends to the use of any and all detection means referred to *supra* for the purposes of diagnosing *Plasmodium* infection in humans and other  
15 animals. Variations of the embodiments described herein are described in detail by McPherson *et al.* (1991), which is incorporated in the references.

A further aspect of the present invention contemplates a kit for convenient detection of a *Plasmodium ssp.* in a biological sample.  
20

In an alternative embodiment, the kit of the present invention is also useful for convenient assay of infection by a *Plasmodium ssp.* parasite, wherein the sample being tested is derived from a human or other animal suspected of being infected with said parasite.

25 The kit of the present invention is compartmentalized to contain in a first compartment, one or more nucleic acid molecules which comprise a sequence of nucleotides corresponding to a *Plasmodium* extrachromosomal genetic element or a complementary nucleotide sequence or a homologue, analogue or derivative thereof as hereinbefore defined.

30 In a particularly preferred embodiment, the first compartment is adapted to contain one

or more nucleic acid molecules which are at least 95% identical to the nucleotide sequence set forth in any one or more of SEQ ID Nos:1-4 or its complement or a derivative, homologue or analogue thereof.

- 5        The kit optionally comprises several second containers comprising a reaction buffer suitable for use in one or more of the detection means described herein and optionally several third containers comprising a nucleic acid molecule positive standard, to which the assay sample result may be compared.
- 10       In an exemplified use of the subject kit, a negative control reaction is carried out in which the contents of the first container are contacted with the contents of the second container. At the same time, the sample to be tested is contacted with the contents of the first and second containers for a time and under conditions sufficient for hybridisation to occur. If the reagents contained in the first container provided is not labelled with a reporter molecule, then the
- 15 contents of the first container may be so labeled prior to the hybridisation reaction being carried out. The hybridised test sample and the negative control sample are then subjected to a detecting means as hereinbefore described. In analysing the results obtained using said kit, the control negative control reaction, test sample and nucleic acid molecule positive standard are compared side-by-side. The contents of the third container should always provide a positive
- 20 result upon which to compare the results obtained for the negative control and test sample. If the results of the test sample are identical to the results obtained for the negative control, then the biological sample does not contain a *Plasmodium ssp.* extrachromosomal genetic element. However, if the test sample produces a nucleic acid molecule which is similar or the same as that contained in the positive standard, albeit of different intensity, then the biological sample
- 25 contains a *Plasmodium ssp.* extrachromosomal genetic element.

A further aspect of the invention provides a specific extrachromosomal genetic element probe, derived from *Plasmodium ssp.*, or a homologue, analogue or derivative thereof, according to the embodiments described herein.



In a preferred embodiment, the extrachromosomal genetic element probe is derived from a species of *Plasmodium* other than *P. falciparum*. More preferably, the extrachromosomal genetic element probe is derived from a *Plasmodium ssp.* selected from the list comprising *P. vivax*, *P. malariae*, *P. ovale*, *P. cynomolgi*, *P. gonderi*, *P.*  
5 (*Hepatocytis*) *kochi*, *P. inui*, *P. knowlesi*, *P. reichenowi*, *P. rodhaini*, *P. schwetzi*, *P. cathemerium*, *P. elongatum*, *P. relictum*, *P. lophurae*, *P. gallinaceum*, *P. yoelii*, or *P. berghei*, amongst others.

In a particularly preferred embodiment, the extrachromosomal genetic element probe  
10 is derived from *P. berghei*. More particularly, the extrachromosomal genetic element probe comprises a sequence of nucleotides which is at least 95% identical to the sequence set forth in any one or more of SEQ ID Nos:1-10, or a complementary nucleotide sequence, homologue, analogue or derivative thereof which is at least useful as a probe for the diagnosis of infection of a human or animal subject by a *Plasmodium ssp.*

15

The present invention is further described by the following non-limiting Figures and Examples.

In the Figures:

**Figure 1** is a copy of a photographic representation of an electron micrograph of the *Plasmodium berghei* extrachromosomal plasmid preparation. P denotes the *P.berghei* plasmid while M is pBR322 (4.36 kb) used as a size marker. The bar represent the size of 1 kb.

**Figure 2** is a copy of a photographic representation showing an *EcoRI* restriction digest of *Plasmodium berghei* extrachromosomal plasmid DNA. In panel (a), plasmid DNA was digested with 20 units of *EcoR* I (New England Biolabs (NEB), Beverly, MA, USA ) in a reaction mixture of 10 uL. The digested products were separated on a 0.4 % (w/v) agarose gel at 120 V for 6 hours. Lane 1 shows Lambda Monocut markers (NEB, USA); lane 2 the *EcoRI* digest resulting in three fragments (E1, E2 and E3); lane 3 shows Lambda DNA-Hind III digest markers (NEB, USA); and lane 4 shows Lambda DNA-*BstEII* digest markers (NEB, USA). Panel (b) shows a *HindIII* digest of the extrachromosomal element of *P.berghei*. The digested products were separated on a 0.6 % (w/v) agarose gel at 100 V for 6 hours. Lane 1 shows Lambda DNA-*BstEII* digest markers (NEB, USA); lane 2 shows a *HindIII* digest of *P. berghei* DNA resulting in six fragments (H1, H2, H3, H4, H5 and H6); and lane 3 shows Lambda DNA-*BstEII* digest markers (NEB, USA).

**Figure 3** is a copy of a photographic representation of a Southern Hybridization of *HindIII* and *HindIII* /*EcoRI* digests of the *Plasmodium berghei* extrachromosomal element. Panel (a) shows restriction digests of *P. berghei* extrachromosomal DNA. Lane M1 shows the 123 bp DNA marker (Gibco-BRL); lane HE the *HindIII* /*EcoRI* digest resulting in 8 fragments (H2, H3, H4, HE1, H5, HE2, E1 and E2); lane H the *HindIII* digest resulting in 6 fragments, H2, H3, H4, H5 and H6); lane M2 the Lambda DNA-*HindIII* digest markers (NEB, USA); and lane M3 the Lambda DNA-*BstEII* digest markers (NEB, USA). Panel (b) shows a Southern hybridization of the fragments in panel (a) with probe PS 1. Panel (c) shows a Southern hybridization of the fragments in panel (a) with probe PL. Panel (d) shows a Southern hybridization of the fragments in panel (a) with probe PSL.

- 24 -

**Figure 4** is a representation of the physical and genetic map of the *Plasmodium berghei* circle. Panel (a) is a schematic representation of the arrangement of various genes and the *EcoRI* and *HindIII* sites are shown. The three *EcoRI* fragments, E1, E2 and E3 as well as the *HindIII* fragments H1, H2a, H2b as well as H4 are shown. Fragment H6  
5 comprises of HE1, E1, E2 and HE2. The relative position of the various PCR products (Table 1) is also indicated. Panel (b) shows a comparison between homologous genes on the *Plasmodium falciparum* and *Plasmodium berghei* plasmid circles.

**Figure 5** is a copy of a photographic representation showing RT-PCR analysis of  
10 rRNA transcripts. Lane 1 shows the 123bp DNA ladder (Gibco BRL); lanes 2 and 3 show the RT-PCR products (L and S) using a set of *Lsu-rRNA* gene specific primers and a set of *ssu-rRNA* gene specific primers respectively.

- 25 -

### EXAMPLE 1

#### Preparation of *Plasmodium berghei* extrachromosomal DNA

5        *Plasmodium berghei* (ANKA strain) was maintained in Swiss White mice by continuous blood passage. Development of parasitemia was monitored daily by thin blood film analysis (Shute, 1988). Parasites were obtained by lysis of infected red blood cells with 1 % saponin. The extrachromosomal element was purified from the parasites using a modified procedure of the Qiagen plasmid mini preparation kit (Qiagen Inc., Chatsworths, CA, USA).  
10 Parasites from 10 infected mice (20-25 g) with a parasitemia of 60% were resuspended in 5 ml of P1 buffer, lysed with 5 ml of P2 buffer and neutralised with 5 ml of P3 buffer. After chilling on ice for 20 minutes, the precipitate was removed by centrifugation according to the manufacturer's recommendation and 200  $\mu$ l of proteinase K (50 mg/ml) were added to the supernatant, which was then reincubated for 2 hours at 37 C. The supernatant was  
15 subsequently passed through a tip-20 Qiagen column which had previously been equilibrated with 1 ml of QBT buffer. The column was washed four times with 1 ml of QC buffer each. Finally, the extrachromosomal element was eluted with 1 ml of QF - buffer which was preheated to 65 C. The DNA was precipitated with isopropanol, washed with 70% ethanol, dried and dissolved in 25  $\mu$ L of TE buffer.

20

### EXAMPLE 2

#### Preparation and restriction digest of *P. berghei* extrachromosomal DNA

25        *Plasmodium berghei* extrachromosomal DNA was extracted from the parasite using the Qiagen plasmid mini preparation kit (Qiagen Inc., Chatsworths, CA, USA). Electron microscopic analysis of this preparation showed circular DNA elements of about 10 times the size of control pBR 322 plasmids (Figure 1). The preparation was not homogenous and, in addition to the circular elements many linear molecules of different lengths were observed.  
30 The preparation is enriched for the extrachromosomal DNA elements of both circular and

- 26 -

linear DNA representing the homologues of the 35 kb circle and 6 kb mitochondrial DNA. These are likely to be 6 kb DNA molecules which are tandemly arrayed in head-to-tail configurations.

5       The extracted plasmid DNA was digested into 3 fragments of 15 kb (E3), 10 kb (E2) and 5.7 kb (E1) by *EcoR* I (Fig. 2a). This gives the plasmid an estimated size of 31 kb. *Hind* III digest of the DNA yielded 6 fragments of 22.4 kb (H6), 4.4 kb (H5), 1.85 kb (H4), 1.23 kb (H3), 0.95 kb (H2) and 0.7 kb (H1), respectively (Fig. 2b).

10       The estimated size of the *P. berghei* circle is 31 kb according to Electron microscopic measurements using pBR322 as control as well as from size calculations using the *EcoR* I restriction digest fragments.

Extrachromosomal circular DNA has not only been found in *Plasmodium* species but  
15 also in other parasitic protozoa such as *Babesia* and *Entamoeba* (Gozar and Bagnara, 1995; Egea and Lang-Unnasch, 1995; Sehgal et al., 1994) suggesting a common evolutionary origin of this circular DNA material (Williamson et al. 1994). By maintaining such extrachromosomal information during evolution it appears that this highly conserved and seemingly functional extrachromosomal DNA molecule is important for parasite development  
20 and that knowledge of its functions will greatly aid in providing novel targets for drug development.

Our preliminary tests using an antisense oligonucleotide approach indicate that this extrachromosomal element may indeed be crucial for parasite survival.

25

### EXAMPLE 3

#### PCR amplification and sequence analysis of plasmid DNA.

In order to obtain a genetic map of the *Plasmodium berghei* extrachromosomal  
30 plasmid, polymerase chain reaction (PCR) amplifications and sequence analysis of plasmid

DNA were carried out.

PCR amplifications of different parts of the extrachromosomal plasmid were performed using primer sets homologous to sequences from the 35 kb circle of *P. falciparum* (Table 1).

PCR was performed using the United States Biochemical (Amersham) PCR kit in a 100  $\mu$ l reaction mixture containing 2 mM  $MgCl_2$ , 0.2 mM of each dNTP, 4 ng/ $\mu$ L of each primer, 5 units of *Taq* DNA polymerase, 10  $\mu$ L of the 10 X PCR buffer and 1  $\mu$ L of the extrachromosomal DNA prepared as described in Examples 1 and 2. A "hot start" was carried out at 95°C for 5 minutes without the dNTPs and *Taq* polymerase. This was followed by the addition of dNTPs and *Taq* polymerase and 40 cycles of denaturation (90°C, 1.5 minutes), annealing (55°C, 3 minutes) and extension (72°C, 5 minutes). A final extension was performed at 72°C for 10 minutes.

15

The PCR products were loaded onto a 1 % low melting point agarose gel, extracted by the freeze-thaw method (Shoemaker and Slayers, 1991) and then cloned into the Promega pGEM-T vector.

20 Clone H2a was constructed by cloning the second fragment of a *Hind*III digest of the extrachromosomal element into the pBluescript vector. (Stratagene, USA).

The clones were sequenced using the ABI PRISM Dye terminator cycle sequencing kit from Perkin-Elmer on the 373A DNA sequencer from Applied Biosystems. The percentage homology with the *P. falciparum* extrachromosomal element (Accession No. X 95275 and X 95276) was obtained using the Martinez/Needleman-Wunsch DNA alignment programme from DNASTAR.

25

**Table 1. Description of clones of various segments from the extrachromosomal element in *P. berghei* and their percentage homology with *P. falciparum*.**

Name 5 of clone	Description	Size (bp)	Percentage homology with <i>P. falciparum</i>	<i>EcoRI/HindIII</i> sites
H2a	Second fragment of <i>HindIII</i> digest containing SSU rRNA	949	92.3	<i>HindIII</i> 1, 944
10 PS1	PCR product of SSU rRNA	526	94.3	Nil
PL.	PCR product of LSU rRNA	595	95.5	<i>HindIII</i> 113
PL2	PCR product of LSU rRNA	554	92.7	<i>HindIII</i> 575
PL3	PCR product of LSU rRNA	735	88.8	<i>HindIII</i> 177
15 PLS	PCR product of tRNAs between LSU and SSU rRNA	973	87.3	Nil
PPH	PCR product of tRNAs before the repeat	1000	78.0	<i>EcoRI</i> 105
PLH	PCR product from LSU rRNA to HtRNA	1118	82.3	Nil
20 PL470	PCR product from LSU to ORF470	1125	83.0	Nil
PRB	PCR product of the RpoB gene	516	87.9	<i>EcoRI</i> 273
25 PWQ	PCR product of the Phe and Gly tRNAs	161	69.6	Nil

- 29 -

Amplifications were also performed using the primers set forth in SEQ ID Nos:5-10. In particular, primer sets comprising SEQ ID Nos:5 and 6 (L/L Primer pair), SEQ ID Nos:7 and 8 (L/S primer pair) and SEQ ID Nos:9 and 10 (S/S Primer pair, homologous to the small-subunit (ssu) *-rRNA* of *P. falciparum*) were used.

5

The amplified regions obtained with these primers lay within the large subunit (*lsu*) *-rRNA* gene, *rpo B* gene, the cluster of 10 *tRNAs*, part of the cluster of four *tRNAs* located close to the 3' end of the *tufA* gene in *P. falciparum* as well as the region between the *lsu-rRNA* and the *ssu-rRNA* genes.

10

All PCR fragments were cloned into the pGem-T vector from Promega. Sequence analysis performed using the Martinez/Needleman-Wunsch DNA alignment on all clones which had been purified using Qiagen midi plasmid preparation columns showed a similarity index of greater than 80% with the *P.falciparum* circle except for the PPH and PWQ  
15 fragment (Table 1). The PRB fragment was homologous to the *P.falciparum rpo B* gene with a similarity index of 87.9% for the DNA sequence and 85.6% for the corresponding amino acid sequence (using the Lipman-Pearson protein alignment). The PPH sequence spanning the cluster of 10 *tRNA* genes had a similarity index of only 78%. While the *tRNA* coding regions were highly similar to those in *P.falciparum* the non-coding spaces were much less conserved  
20 between the two Plasmodium species.

In order to examine if the *lsu-rRNA* gene in *P. berghei* exists as a repeat, a single forward primer (L3) homologous to the 3' end of the *P.falciparum lsu-rRNA* sequence and 2 distinct reverse primers homologous to the ORF 470 (04) and the start of the cluster of 10  
25 *tRNA* genes (3H) of the *P.falciparum* circle were designed. The fragment amplified with the L3/04 primer set (PL470) was distinct from that amplified using the L3/3H set (PLH). Sequence analysis of PL470 showed a homology of 83% with the same region in *P. falciparum*. The sequence of PLH was homologous to the 3' end of the *lsu-rRNA* and the 3' end of *rps 4* of the *P.falciparum* circle (data not shown) indicating that the *lsu-rRNA* gene  
30 exists as a repeat in *P.berghei*. In addition, a cluster of intervening *tRNA* genes was present



- 30 -

between the *lsu-rRNA* and *ssu-rRNA* genes (fragment PLS). This repetition and arrangement is similar to the organisation of the *P.falciparum* circle, where a palindromic sequence of genes for the subunit *rRNAs* and several *tRNAs* exists. Each arm of the palindrome contains one *ssu* and one *lsu-rRNA* gene and a cluster of intervening *tRNA* genes (Gardner *et al.*,  
5 1993).

From the nucleotide sequences set forth in SEQ ID NOs:1-4, it is clear that the genes in the *P.berghei* circle are homologous to those in the 35 kb *P.falciparum* circle. Major differences in sequence are observed in the non-coding spaces between tRNA gene clusters.  
10 The arrangement of genes appears to be similar in both Plasmodium species and a repeat of the rRNA genes does not only exist in *P.falciparum* but also in the *P.berghei* circle.

15

#### EXAMPLE 4

##### Southern Hybridisation of restriction fragments

Fragments from the *Hind*III and *Hind*III/ *Eco*RI digests of the *Plasmodium berghei* extrachromosomal element were separated on a 1 % (w/v) agarose gel at 120 V for 4 hours.  
20 The separated fragments were then transferred onto a Nylon membrane (Hybond-N, Amersham) by capillary action using 20x SSC buffer (0.3M sodium citrate, 3M sodium chloride, pH 7.0). Southern hybridization was performed using probes that were made from the cloned PCR products, according to the preceding Examples.

25 The PCR products were liberated from the vector by digestion with *Apa*I and *Pst*I. The enhanced chemiluminescence (ECL) direct nucleic acid labelling and detection system (Amersham International PLC, England) was used for labelling the probe, for hybridisation and for detection.

30

Each of these steps were carried out according to the manufacturer's instruction. First, 8  $\mu\text{g}$  of probe in a volume of 20  $\mu\text{L}$  were denatured by boiling for 5 minutes and immediately cooled on ice for 5 minutes. 20  $\mu\text{L}$  of labelling reagent were then added. This was followed by the addition of 20  $\mu\text{L}$  of glutaraldehyde solution. The mixture was incubated for 20 minutes at 37°C before addition to the hybridization buffer. The ECL Gold hybridisation buffer containing 0.5M NaCl and 5% blocking agent was used for hybridisation. The blots were prehybridised for 2 hours at 42°C and the labelled probe was added to a final concentration of 800 ng DNA/ml. Hybridisation was allowed to proceed overnight at 42°C. The blots were washed twice in primary wash buffer containing 6M urea, 0.4% SDS and 0.5x SSC at 42°C for 20 minutes. This was followed by two rounds of washing in 2x SSC buffer at room temperature for 10 minutes. For detection, 6.5 ml of equal volumes of detection reagents 1 and 2 were mixed and added to the blot for 1 minute. The blot was then drained, wrapped in Saran Wrap and the DNA side was exposed to an autoradiography film.

15 A double digest of the *P. berghei* circle with *Hind*III followed by *Eco*RI resulted in the following fragments: 10kb (E2), 5.7kb (E1), 5.0kb (HE2), 4.4kb (H5), 2.3kb (HE1), 1.85 (H4), 1.23 (H3), 0.95(H2) and 0.7kb (H1). The PS 1 probe hybridised to H2, the PL probe hybridised to H6 and HE2 while the PWQ probe hybridised to H6 and H2 (Fig. 3).

20 The results obtained with various other probes are shown in Table 2. Of interest to note is that H2 contained 2 distinct fragments which hybridised with PS 1 and PL3. One of the H2 fragments (H2a) was cloned into Bluescript vector (pBS KS (II)+) and sequenced. The sequence corresponded to the internal region of the *P.falciparum* 35 kb ssu-rRNA gene (Table 1, sequence H2a). The other fragment (H2b) arose from the two *Hind* III sites within  
25 the lsu-rRNA gene. The presence of these two sites was confirmed by the sequences from the PL2 and PL3 PCR products. In addition, both the H5 and the H3 fragments hybridised with probes corresponding to the *P.berghei* 6 kb mitochondrial DNA. This was not unexpected as the preparation was found to contain linear molecules as shown by electron microscopy (Fig. 1).

Table 2. Southern analysis of restriction digests.

5	Probe	<i>Hind</i> III digest	<i>Hind</i> III/ <i>Eco</i> RI digest	<i>Eco</i> RI digest
10	PS1	H2	H2	N.D.
	PLS	H4	H4	N.D.
	PWQ	H6	E2	E2
15	PL.	H6	HE2	N.D.
	PL2	H6	HE2	N.D.
20	PL3	H2, H4	H2, H4	E3
	PRB	H6	HE2	E2, E3
	PL470	H6	HE2	E3
25	PPH	H6	HE1	E3
	PbIV2.3	H3, H5	H3, H5	N.D.
30				

### EXAMPLE 5

#### Physical and genetic map

A map of the *P. berghei* extrachromosomal circle was constructed based on the information from the restriction digests, Southern hybridisation experiments and the sequence analysis of the PCR fragments (Fig. 4a). The PPH and PRB fragments each contained an *EcoR* I restriction site (Table 1). The three *EcoR* I and six *Hind* III fragments were arranged according to their hybridisation patterns. The PRB probe hybridised to both the E2 and E3 fragments from the *EcoR* I digest indicating that E2 is positioned next to E3 (Table 2).

10

The HE2 fragment obtained from the double digest with *EcoR* I and *Hind* III, hybridised to probes PRB, PL470 and PL2 while H2b and H4 hybridised to probe PL3. Both the PL2 and PL3 fragments are regions within the *lsu-rRNA* gene, whereas the PL470 fragment contains 3' end of the *lsu-rRNA* gene. Thus, the ORF470 must be located next to the PL2 fragment. H2b is situated between HE2 and H4 since H4 also hybridised with the PLS probe which contains the 5' ends of both the small and large subunits rRNA genes. H2a hybridised with probe PS 1 which corresponds to a region within the *ssu-rRNA* gene, therefore H2a must be located next to H4. Finally, HE1 is placed next to E1 as HE1 hybridised to probes PPH, PLH and PL2.

20

The arrangement of genes on the *P. berghei* circle spanning the *rpo B* gene and the cluster of 10 *tRNAs* genes is thus very similar to that of the *P. falciparum* 35 kb circle (Fig. 4b). The *P. berghei* circle encodes organelle-like rRNAs, tRNAs, ribosomal proteins and RNA polymerase subunits, similar to those identified for *P. falciparum* (Preiser *et al.*, 1995).

25

- 34 -

### EXAMPLE 6

#### Reverse transcription-PCR of LSU-rRNA and SSU-rRNA

In order to determine if the *Plasmodium berghei* extrachromosomal genetic element  
5 is transcriptionally active, total RNA from *P. berghei* was isolated using the RNeasy total  
RNA kit (Qiagen Inc., Chatsworth, CA, USA) and a combined reverse transcription - PCR  
(RT-PCR) reaction was carried out to amplify *lsu-rRNA* or *ssu-rRNA* transcripts.

Total RNA was isolated from *Plasmodium berghei* using the Qiagen RNeasy Total  
10 RNA kit (Qiagen Inc., Chatsworth, CA, USA). Parasites from 10 infected mice with a  
parasitemia of 60% were resuspended in 350  $\mu$ l of lysis buffer RLT and homogenised using  
a QIAshredder (Qiagen Inc.). The homogenate was cleared of insoluble material by  
centrifugation and 1 volume of 70% ethanol was added. The entire sample was then added  
to the RNeasy spin column and washed with RW1 buffer followed by two washes with RPE  
15 buffer. The RNA was eluted out with 35  $\mu$ l of water. 5  $\mu$ l of the RNA was used as starting  
material for the Access RT-PCR system (Promega, Madison, USA). Two primer sets, L/L  
(SEQ ID Nos:5 and 6) and S/S (SEQ ID Nos:9 and 10) were used. The manufacturer's  
protocol was followed with the exception of the annealing step for PCR amplification.  
Annealing was allowed to proceed at 55°C for 1 minute. The PCR products were separated  
20 on a 1% (w/v) agarose gel and visualised by ethidium bromide staining.

Amplification using the RT- PCR kit from Promega and a set of primers homologous  
to the *ssu-rRNA* produced a 526 bp fragment while amplification using a set of *lsu-rRNA*  
specific primers resulted in a 600bp fragment (Fig. 5).

## EXAMPLE 7

Assay of blood samples for the presence of *Plasmodium ssp.*

A total of 102 *Plasmodium*-infected blood samples from four different locations,  
 5 Singapore, Laos, Karachi and Calcutta, and a defined number of negative control blood  
 samples, were analysed for the presence of *Plasmodium* extrachromosomal genetic elements,  
 using the polymerase chain reaction.

Briefly, 10-100 $\mu$ L of whole patient blood (either peripheral blood from a finger prick  
 10 sample or venal blood) was spotted onto a filter disc or equivalent solid support and directly  
 amplified, using each of the primer pairs:

## L/L PRIMER PAIR:

SEQ ID NO:5: 5'-GACCTGCATGAAAGATG-3'  
 15 SEQ ID NO:6: 5'-GTATCGCTTTAATAGGCG-3'

## L/S PRIMER PAIR:

SEQ ID NO:7: 5'-GCCACTACTATGAAAATC-3'  
 20 SEQ ID NO:8: 5'-GCGTTCATTCTGAGCTAG-3'

## S/S PRIMER PAIR:

SEQ ID NO:9: 5'-GCGGTAATACAGAAAATGCAAGCG-3'  
 SEQ ID NO:10: 5'-AGCACGAACTGACGACAGCCATGCAC-3'

25 PCR Buffer used in the amplification reactions comprised the following:

70mM Tris. pH8.8  
 20mM Ammonium sulphate  
 1 mM DTT  
 0.1 $\mu$ g/ $\mu$ L BSA  
 30 2.5mM MgCl<sub>2</sub>

- 36 -

Each 100  $\mu$ L reaction included 0.4 $\mu$ g of each primer, 0.8mM dNTP mixture and 5U of *TaqI* polymerase.

5        The template DNA was fixed with methanol for 5 mins. A "hot start" was carried out at 95°C for 5 minutes without the dNTPs and Taq polymerase. This was followed by the addition of dNTPs and Taq polymerase and 40 cycles of denaturation (90°C, 1.0 mins), annealing (60°C, 2 mins) and extension (72°C, 3 mins). The PCR products were analysed by agarose gel electrophoresis.

10

The results are shown in Tables 3 and 4. The L/L primer set was capable of identifying *Plasmodium falciparum*, *P. vivax* and *P. malariae* in 100% of cases, suggesting that this primer pair is useful in the genera-specific diagnosis of *Plasmodium* infection. The S/S primer set was capable of efficiently diagnosing *P. falciparum* and *P. malariae* in 100% of cases.

15 In marked contrast, the L/S primer set resulted in only poor diagnosis of *P. vivax* and *P. malariae*, however detected the presence of *P. falciparum* in blood samples, suggesting that this primer pair is species-specific.

Results also indicate that the selection of primer pairs in the diagnostic assay was of  
20 primary importance in determining the reliability of the assay in diagnosing infection by *Plasmodium ssp.*

- 37 -

**TABLE 3**  
**Number and origin of *Plasmodium*-infected blood samples**

ORIGIN	TOTAL No. OF SAMPLES	<i>Plasmodium</i> <i>falciparum</i>	<i>Plasmodium</i> <i>vivax</i>	<i>Plasmodium</i> <i>malariae</i>
Singapore	13	7	5	1
Laos	11	10	1	0
Karachi	67	14	53	0
Calcutta	11	1	10	0
<b>TOTAL</b>	102	32	69	1

**TABLE 4**  
**PCR results using the primer pairs L/L, L/S and S/S**

<i>Plasmodium</i> ssp.	L/L Primers	L/S Primers	S/S Primers
<i>P. falciparum</i>	32/32 (100%)	14/20 (70%)	21/21 (100%)
<i>P. vivax</i>	69/69 (100%)	6/58 (10%)	48/57 (84%)
<i>P. malariae</i>	1/1 (100%)	0/1 (0%)	1/1 (100%)
<b>TOTAL</b>	102/102 (100%)	20/79 (25.3%)	70/79 (88.6%)

Those skilled in the art will appreciate that the invention described herein is susceptible to variations and modifications other than those specifically described. It is to be understood that the invention includes all such variations and modifications. The invention also includes all of the steps, features, compositions and compounds referred to or indicated in this specification, individually or collectively, and any and all combinations of any two or more of said steps or features.



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- 40 -

SEQUENCE LISTING

(1) GENERAL INFORMATION:

5

(i) APPLICANT: THE NATIONAL UNIVERSITY OF SINGAPORE

(ii) TITLE OF INVENTION: DIAGNOSIS OF PARASITES

10

(iii) NUMBER OF SEQUENCES: 10

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(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk  
(B) COMPUTER: IBM PC compatible  
(C) OPERATING SYSTEM: PC-DOS/MS-DOS  
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25

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(A) APPLICATION NUMBER: AU PROVISIONAL  
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30

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(2) INFORMATION FOR SEQ ID NO:1:

40

(i) SEQUENCE CHARACTERISTICS:

- 41 -

(A) LENGTH: 5507 base pairs

(B) TYPE: nucleic acid

5 (ii) MOLECULE TYPE: DNA

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Plasmodium berghei

10 (vii) IMMEDIATE SOURCE:

(B) CLONE: CLONE PSI-L470

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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 TAATGAAGCT GTACATCCTT CTAAATATCC AACATATGCA AATTCACCTG CTATTAATAA 60  
 AGTACGTTCA AATTGTGCAA AATCATAAGA ATTAGTCTTA AAATAAGTTG ATAAATTAAA 120  
 20 ACTACATTTT ATATACTTAG ACACATAACA AAAAGATCCT TCACTAAAAA TAATTGAATT 180  
 AATATTTGCA AAAAAATTAT CTTATAAGA AACTACAGT TCCTAAATAT TTTTITACTA 240  
 ATAAAGGATA TTTTAAAATA ACGTCCAATA AGAGACAAAA ATATAATACC TAATTTTTTT 300  
 25 AAAAAATATT GTGTTGTATG TTAAACAGA TATACTATCC AAAAAATAACA TCAAAGGAAT 360  
 TATTTTTTTA TTAAATAGG TATCTAAAAA ATTTATATTT AAATTAGTTT TTAAATATAC 420  
 30 TCACAAATTA CTATCTTTTG AGGGAGGAGA ATAATAAATA ATATTATCAT AACTAATATT 480  
 GGGACATTCTG AAACACGACC AATCTGGTAA TTTACACATA TTAAAAAATT TTAGAGAATA 540  
 TATTTTAAAT TTGTAAATAA AAAAATATAA ATAAATATTA TGAGATAAAT TTTTCTCAA 600  
 35 ATTTTATTTT CTCCATTTCT TATTAAATAT AAATTTATTT TATTATTATA TTGATATTTA 660  
 TAATTTAAAT TATAAATATT TAAAAATTTT TTTAATTTTA ATTTATTTAT CATAATAATT 720  
 40 TTATATTATA AAATATTTCA AGTTAACGAT GAGATTTGAA CTCACAATCT ACTGATTACA 780

- 42 -

	AATCAGTTGC TTTACCAATT AAGCCACTTT AACAAATATA ATATTTATAA TTAAATATTC	840
	AACTTATTAG GAATTATACA CAAAATATAT TACTATAAAT ACATATTAAT TCTATAAAAT	900
5	AATTTTCTA ATTATTGTTT TATTCATTTA TATGATTAGA ATATTATTTT TAATTAAATT	960
	TTCTTATTTA TATTACTTCA ACAATTAAAA TTTTATACTT AACTACTCAA CATTACAAAA	1020
	TATAATAATT GATATATCAT TGGTATAATT TTTTCGATCC TCTCGTACTA GAAAAAATAA	1080
10	TTCAATATT CTAACACTTA TATTAGATAT GGACCGAACT GTCTCACGAC GTTCTGAACC	1140
	CAGCTCACGT ATCGCTTTAA TAGGCGAACA GACTTACCCT TAAACATAC TACTGCCTTA	1200
15	GGATGCGATA AGCCGACATC GAGGTGCCAA ACCTTTTCGT CAATATGGAC TCTCGGAAAA	1260
	GATTAGCCTG TTATCCCTAG AGTAACTTTT ATCCGTTAAG CGATAATTTT ATTATTAAAT	1320
	AATTATCGGA TCATTAAGAC CGACATTAAT CTCTGTTTAA TTTGTAAATT TTACAGTTAA	1380
20	TTATATATTT ATCTTTATAT AATAAATATA ACATTGTACA CCTCCGTTTT TATATAGGAG	1440
	GAGACCGCCC CAGTCAAACCT ATCTTATAAA TATTGTTAAA AATTTTGTTA TAAAAATTTT	1500
25	ATAAGAATTT ATATATATAT AAAATGGTAT TTCATTAACA ATTACATTAT TTCCAAAAAA	1560
	ATAATATTAC TACTTCCCAT TTATTCTATG TTATATATAT ATATTTTCAA TATCTATTAA	1620
	TAGTAAAGCT TCATAGGGTC TTTCTGTCCT AATATAAGAA ATCTGCATCT TCACAGATAA	1680
30	TTTTATTTC AAGATTTT TTTAAGACA GCATTTAAGT CGTTACATCT TTCATGCAGG	1740
	TCGGAACCTA CCCGACAAGG AATTTGCTA CCTTTGGACC GTTATAGATA CAGCCGCCGT	1800
35	TTACTATAGC TTATATATAT ATTATAATTT TAAATTATAA ATATTATTTT TACATAATAG	1860
	CACTGGGCAG ATGTCAATCT TTATACATCA TCTTTCGATT TAGCAAAGAT TTGTGTTTTT	1920
	GTAAACAGT CGCTTAAATT TTTGTTTTT AACTAAATAA GTATCTCTTC TCCCCTAAGT	1980
40	TTACGAGATA AATTTGCCGA GTTCCTTAAA AAAAATTATC TCAACTTCTT AATAATTTAT	2040

- 43 -

	ATATATTTAC TAGTGTCAGT TTACAGTACG AATACATAAT AATATATATA TATAAATAAT	2100
	TTTTATATAA TATAATATAT TTATTATTAT ATTAGTTTTA AAATATAAAT ATTATTATAT	2160
5	AGTATAAGAA TATTAACCTA TTACCTATCG ATTACACATT ACATCTCATC TTAAGATTCG	2220
	ACTAACCCCTA TTTAAAATAA TAATAAATAG GAGCCCTTAA ATTATAGAAG TATTGGATTT	2280
	TTACCAATAT TTACATTACT CAAATTAGCA TTATCACTTT TGATTTAATT ATTTTAACTT	2340
10	TTCATATAAA TAATTTATAT TCAAAACGCT CTTTACCAA TTTAATTTA TTAATATTAA	2400
	ATTTTATACA TATCGATAAT TAATTTATTT TCGATTATTT CTGAACTAAA ATTACTAAAT	2460
15	TAATGAGCTG TCACGCACTC TTAAAGGGAG AACTGCTTCT AAATTTACTT TGGGGGATGT	2520
	AAATAATTTT AGATTCTTTT TAGGACTTAA TTAATATCTA GAAATCTTAA TTGAGAACTC	2580
	GGGCTGTTTC CCTTTTGAAA ATAAAGCTTA TCCTTTAGTT CCTGATCATA TATATATTTT	2640
20	ATTAAATAAA ATTCTTAAAT TATTTTCATT AATATTAACT ATATAAATTA ATTTAATAAA	2700
	AAAAGAGTTT TACATTTATT TATATATAAA TACTATACTT ACATATATTT CAAAGAGAAC	2760
25	CAGCTATCTT CAAATTCGAT TGGCATTTC ACTCTAATTA TACTTTATTT GATACTTTTG	2820
	CAACAGTAAC CAATTCAAAC TTCAATTTAA TTTTATTTAA ATCTTATTTT AAATATAATT	2880
	AGATCATTG ATTTCTGGGTC TATAATAAAT AATATACTAA ATGCTTATTA TATATAATAA	2940
30	CAAACCTGAG TATACTTTGG CTTCAATTTAT AAATATTTAA CCTAATAATT ATACTATTTA	3000
	TTATAACTTG CTAATTCCTT CTTCAACAAG AAAATAATAA AATTATATTA AATTTTATTA	3060
35	TTATTTATTA AATTTAAAAT TCAGGTTCTT TTCACTATTT TCTCAAAATC CTTTTCATCT	3120
	TTCCCTCACG GTACTATTCA CTATCAACTT TTATTATATT AAATTTTATA AGATAACTCT	3180
	TAATTATATT TATATTATTC ATATAAAATA TATTTTATA TTACTTAATT AAAATTTTAC	3240
40	ATATATAATG TTTTAAATCT TTCAGTTCGC TCGCCACTAC TATGAAAATC GTTATTACTT	3300

- 44 -

	TATATTCCTT TAAGTACTAA GATGATTCAG TTCCTTAAGT TTTTAAAA TATTATATA	3360
	AAAATAAATT TTTATTCAGA TACTTTTATA ATTTAATAA TAAAAAATT TAAATATATT	3420
5	TAATTTTTTA TAATTATAAA AATTTCGTTA ATATATTTAA CGTCTTCTT CAATAATAAA	3480
	AATAATAGAC ATCCTTTTAA ATTTATTATA TATATTTAAT TATATATTTA ACTATATAAA	3540
10	TTATAAATTA ATTTATTTAA AATAAGCGAA AAACGGAATT GAACCGATTA CCTTCGGAGC	3600
	ATGAATCCGA CGAACTTTCC TTATGCTCTA TTTCGCTAAA TACAATTAAA CTGAAAAGA	3660
	ATTGAACTTT TATTTTATAA TTCGTACTTA TATATTTTAT CCATTAAATT ACAAGTTCAT	3720
15	TATATTATAA TATATAAATT ATAAGTAATT AACTTAGAGG TAAAGTTTCT GCTTTACATA	3780
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	TATGGCGAAA AGGTAAACGC GCTAAATTTA GAATTTAGTT TTTATAATAA TAAGAGTTCG	3960
	AATCTCTTTA TTCATATTTA TAATATACTT CTAAACTAG GATTGAACTA GTATCTTTCG	4020
25	GTAAACAGCC GAATGCTTTA ACCACTAAGC TATTAAGAAT ATTAATATTA TATTATATAA	4080
	TATATAATAG GGAATATAGT TTAATGGTAA AATCTTATTC TTGCATAATA AAGATAGTAG	4140
30	TTCAATTCTA CTTATTTCCA TATTATAAAA TCTATAAATG TTATAATTTT TAAATAATAT	4200
	ATATATAATT ATATTGCGAG TTTGATCCTA GCTCAGAATG AACGCTAGAA ATATACATTA	4260
	CACATGCAAA TTTATGGATT ATATCATAGT GAATAGGTGA GGATATATAA ATTTTAAATT	4320
35	TTAAATAGAT TATAATATAT AATAATCTAT AAGCGCATTT ATTTATATAA TTGTACTATA	4380
	TTAAAAATTA TTATTGTTTA AAATAAAATT TATATTTGAT TAACTAGTTG GTAAAATAAA	4440
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- 45 -

GCTTGATAAA GTAATATTTC TTAAAGGATG ACAGTATATT TTTATATTGT AAACCTTTATA 4620  
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 5 GCAGCGGTAA TACAGAAAAT ACAAGCGTTA TTCACCTTAT TTGGCGTAAA GCGTTTTAAG 4740  
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 25 ATAATATATT TATTAAATAT ATAAAAAAGA CTACGTCAAG TCATTATGCT CCTTATATTT 5340  
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 TTAAATATAT TTATAGTTCT GATTATAAAT TGAAACTCAT TTATATGAAG ATGGAATCAC 5460  
 30 TAGTAATCGC TAATAAGAAG TATAGCGGTG AATAAGTTCT TAAGCTT 5507

35 (2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1705 base pairs

(B) TYPE: nucleic acid

40

(ii) MOLECULE TYPE: DNA



- 46 -

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Plasmodium berghei

(vii) IMMEDIATE SOURCE:

5 (B) CLONE: CLONE PLH-PPH

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

10	CAGAAAAATA GGATTGAAC CTATATTCTT CTATTCCCAA AATAGATATG TTACCATTAC	60
	ACTATATTCT GAATATTTAA AATTTTATAC TTTTAAGGAA AATCGAATTC CTATTTTCTT	120
	CTTGAAAAAA AGATGTCTTA CCTTTAAACG ATAAAAGTAA AACTTAAAT TACCTGAGAC	180
15	TTGAACTCAG AACCATTCTGA TTAAGAGTCG AGTACTCTAC CAATTAAGCT AGTAATTCTT	240
	AATATAACGA ATCTGACGAG AATTGAACTC GTATTCTTTG TTATGACAAA ATAATATTTT	300
20	AACCTAATTA AACTACAAAT TCAATAAATA TATATAGGAA AGGGATTGAA CCCTGGTATA	360
	TATAATATCT ACATAAATTG CAATTTATAG CTATAACCAC TCAGCCATTT CTGTATATAA	420
	TAATAAGTTA AATCAGATTG AACTGATGTA GATATAAAAC CCAATGGATT TACAGTCCAT	480
25	CCCTTTTAAC CCCTCAGGCA TTAACTTTAT TATACATTTA AGTAGATTCG AACTACTGAT	540
	GTTCAATATT TGAAAATGAA TTATGAGTCC ATTGCTTTTC ACCTCTTAGC TATAAATGTT	600
30	TACTTTATTA GAGATAAAGG GACTCGAACC CTTACAACAA TTATTGTTAA TGGATTTTCT	660
	AATTGAATTT AGACTTTTTTA TAAACATGTA TATAAATAAT AAAGTCGTTT GAATATATAA	720
	CTAATATATT ACAGAATAAA AATTATTTTT TCTTTATATA TATTTAAATT ATTAATTTAT	780
35	TTATAAAATT AACTCATAAA CAACGAATAT AAATTATATT TATATTATTT AAAGTCCATT	840
	GTGTATACCA AATTCACCA TATCTCTATT ATATACTATA TAAATGATAT TCAGATTTGA	900
40	ACTGAAATAA AATAATTTGC AATTATCCAC TTTACCTAAT TAAGTTATAT CATTATTATA	960

- 47 -

TATTATAAGA TAAATAAAGA GATTTGAAC T CATATAAAAG AAACCACAAT TCCTTATCTT 1020  
 AACCTTTAGG ATTATATTTA TCATTATTAA AACTTATTAT ATATTATAAA TATTATTATA 1080  
 5 AATATATTAA ATATTATTTA AATATAAATC ATTTAATATT TTTATTTTAA AATTATATAT 1140  
 ACATATAATA AAATTATCAT TAAGAACTAG AAGATTTAAT AAAATTATAT TTATATAAAT 1200  
 TTGATATATA AATATATATA TTATATCTAT AAATTAAATT TTGTGAAATT ATATATTTAA 1260  
 10 TTTTTTTATT AAAAAAATT ATATCCTTAC CTCTTTAATT TAATATTATA ATAATTACCA 1320  
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 15 TTTAAAACC CTATATTAAC TAAAAAGACA TCTAATCTAG ATTCTAATAA ATTTAATAAT 1440  
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 TTAAATGTTA TACAATAATT ATAACAGATA TATCTTACAA ATTTTAATTT TAAATCAAAA 1560  
 20 TAAGATTAT AGGACATATT TATTAATTTT TAATTTATAA TTATATTTAC TAGATAAATA 1620  
 TAATAAAAAA GGAAGATTTA ATTTTTTTAA AATTTTATT TTAGGAGTTA AAAATTTTAT 1680  
 25 CATAATAATT TTATATTATA AAATA 1705

(2) INFORMATION FOR SEQ ID NO:3:

30

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 516 base pairs

(B) TYPE: nucleic acid

35

(ii) MOLECULE TYPE: DNA

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Plasmodium berghei

40

(vii) IMMEDIATE SOURCE:

(B) CLONE: CLONE PRB

- 48 -

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

5 TTAATAGACA TGGACATAAA GGTGTTATTT CTTATATTAA TGATATTAAT GATATGCCTT 60  
 ATTTAAATAA CAAAATACAA CCTGATTAT TGTAAAGTGC TATTGGTATA CCTTCTAGAA 120  
 TAAATATAGG TCAAATATTA GAGGGTATAT ATGGATTAAA TAGTTTATAT TTAAATAATA 180  
 10 GATATATAAT ATCTAATAAT TTAAATACTA ATTATTATAA TAATTATATT AATAATTTTA 240  
 ATTATTATAA ATATAATTAT AATAATAATT TTGAATTCAA TAAAATATCA TATAATTATA 300  
 ATAAATATTT TTAAAAAAT CCGTTTACGG GCCATTTAAT ACAGAATAGT ATTTGTTTAA 360  
 15 ATAATATTTA TTATTATAAA TTAGTACATA TGGTAAAAGA TAAATTAAGA TATAGATTCA 420  
 TAGGATTATA TTCTGAATTA ACTCAACAAC CTGTAAAAGG AAATACAAAA CAAGGAGGTC 480  
 20 AAAGATTGCG TGAAATGGAA GTATGGGCGC TAGAAG 516

(2) INFORMATION FOR SEQ ID NO:4:

25

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 161 base pairs
- (B) TYPE: nucleic acid

30

(ii) MOLECULE TYPE: DNA

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Plasmodium berghei

35

(vii) IMMEDIATE SOURCE:

- (B) CLONE: CLONE PWQ

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

40

GTTCAAAAAT CAGATTTGAC TGATAACACA TGGAAGTTCA ATCCATTGCT CTACCATTGA 60

- 49 -

GCTATAATGA CTTAATAATA TTATTATTAT AATAGAATAT AACCAAAAGG TTAAGGTAAT 120

GAACTTTGAT TTCATTAATA TAGGTTGAA TCCTTTAGGA C 161

5

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- 10 (A) LENGTH: 17 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

15 (ii) MOLECULE TYPE: DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

20

GACCTGCATG AAAGATG

17

25 (2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- 30 (A) LENGTH: 18 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA

35

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

GTATCGCTTT AATAGGCG

40

18

- 50 -

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 18 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

15 GCCACTACTA TGAAAATC

18

(2) INFORMATION FOR SEQ ID NO:8:

20

(i) SEQUENCE CHARACTERISTICS:

- 25 (A) LENGTH: 18 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA

30

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

GCGTTCATTC TGAGCTAG

18

35

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

- 40 (A) LENGTH: 24 base pairs  
(B) TYPE: nucleic acid

- 51 -

(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA

5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

10 GCGGTAATAC AGAAAATGCA AGCG

24

(2) INFORMATION FOR SEQ ID NO:10:

15

(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 26 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

20

(ii) MOLECULE TYPE: DNA

25

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

AGCACGAACT GACGACAGCC ATGCAC

26

30

DATED this SIXTH day of FEBRUARY, 1997

The National University of Singapore

by DAVIES COLLISON CAVE

Patent Attorneys for the Applicants

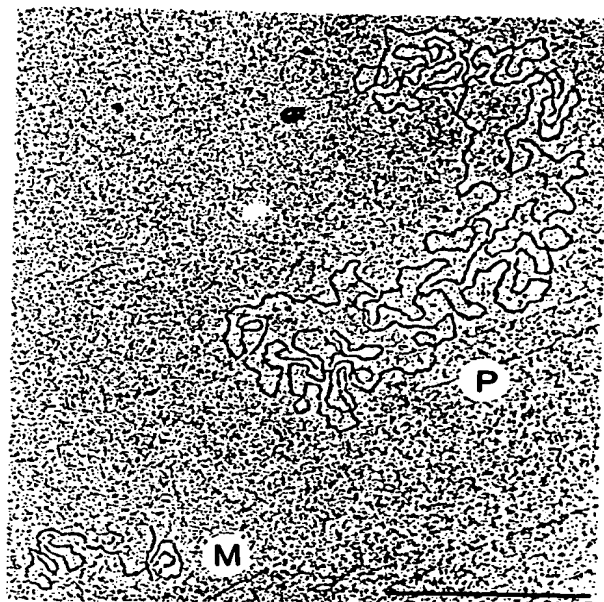


FIGURE 1

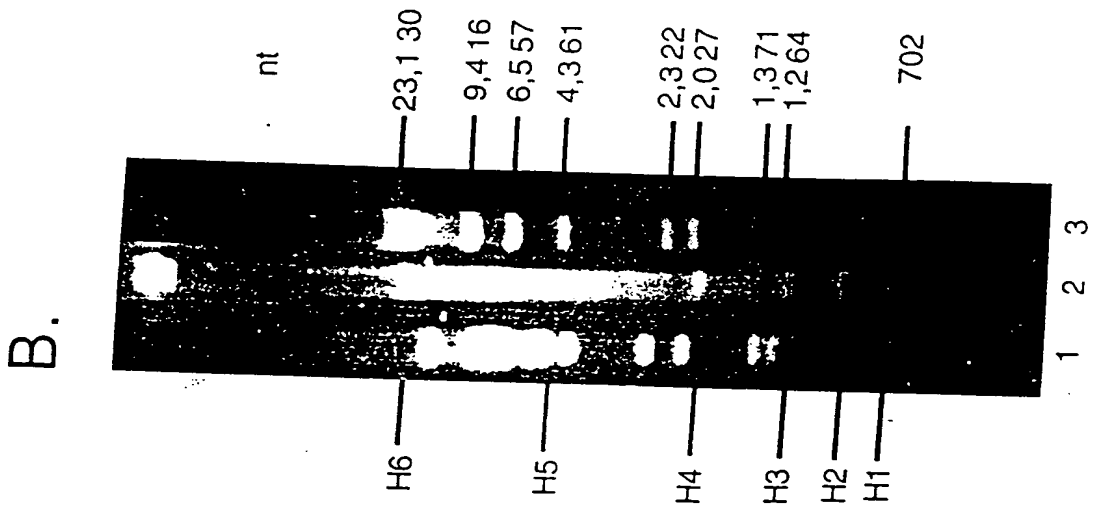
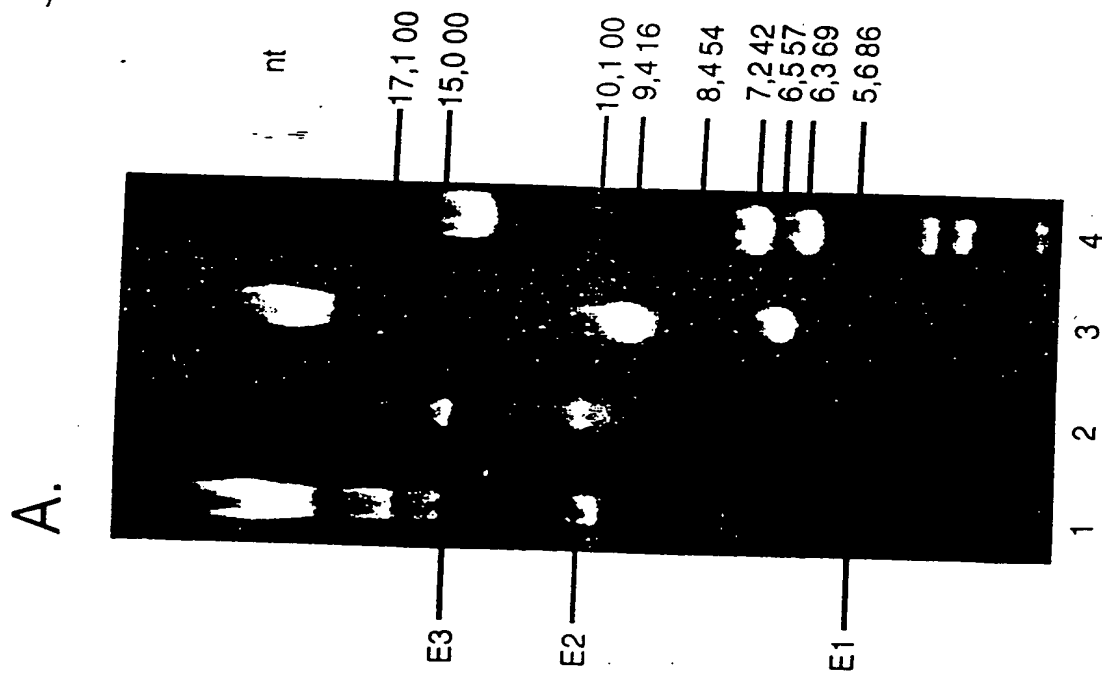


FIGURE 2



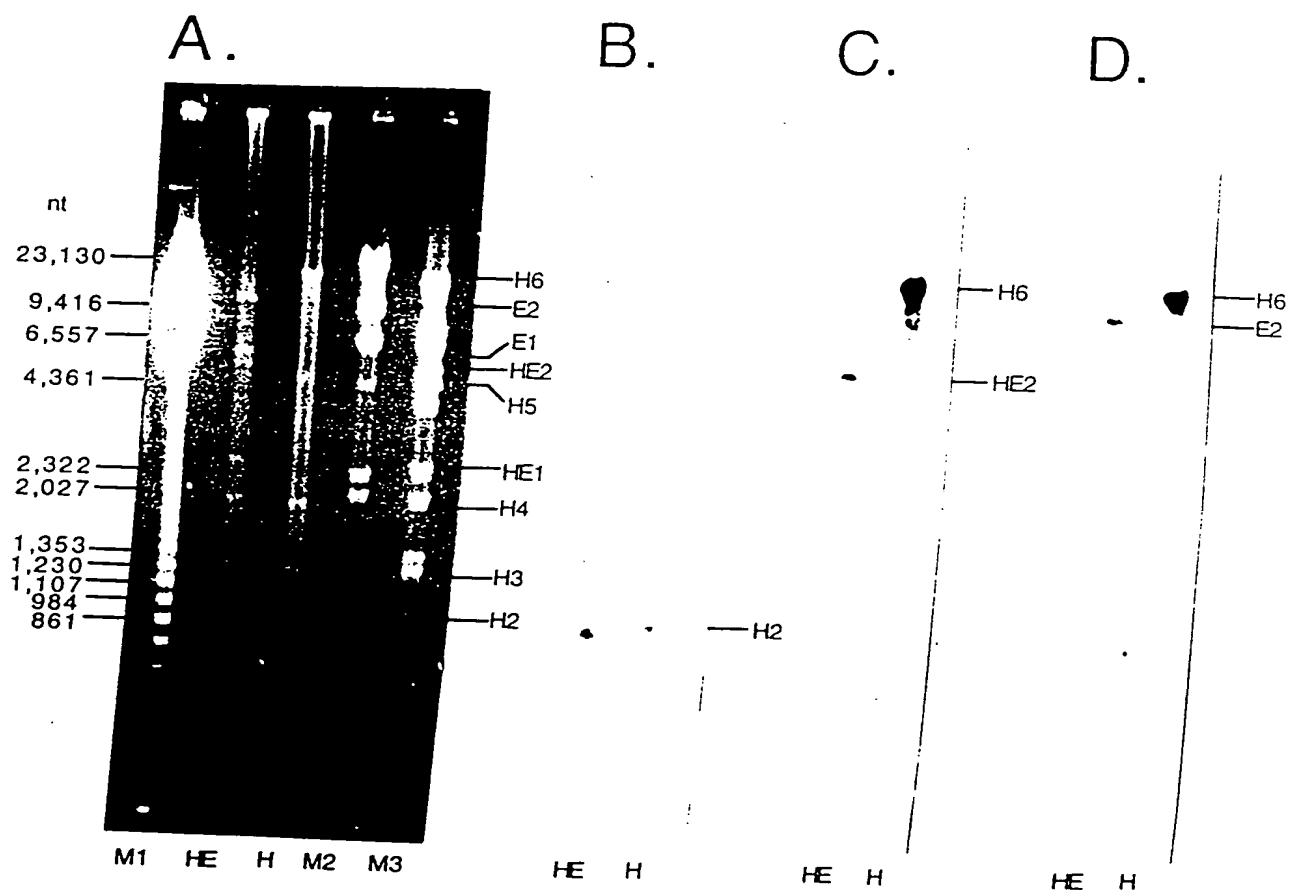


FIGURE 3

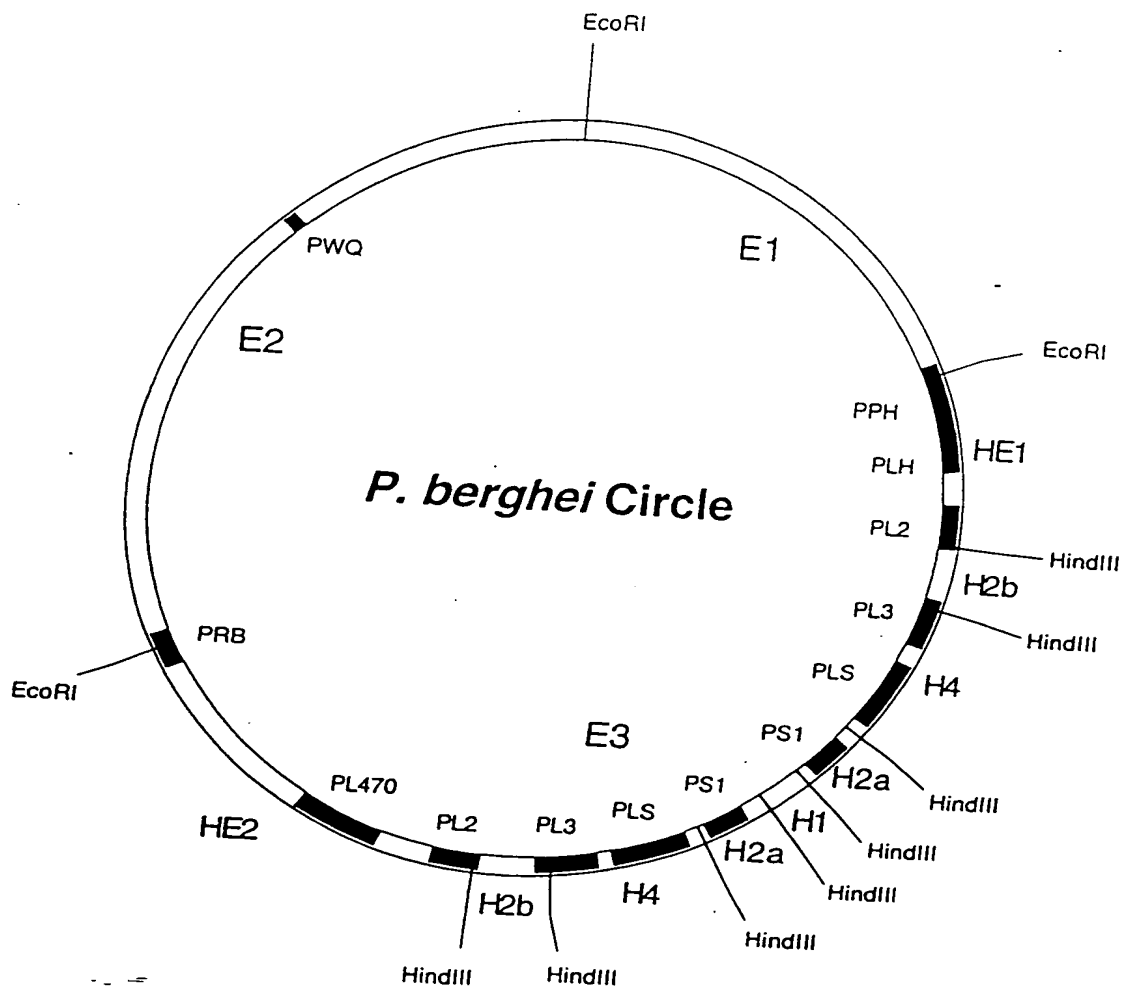


FIGURE 4A

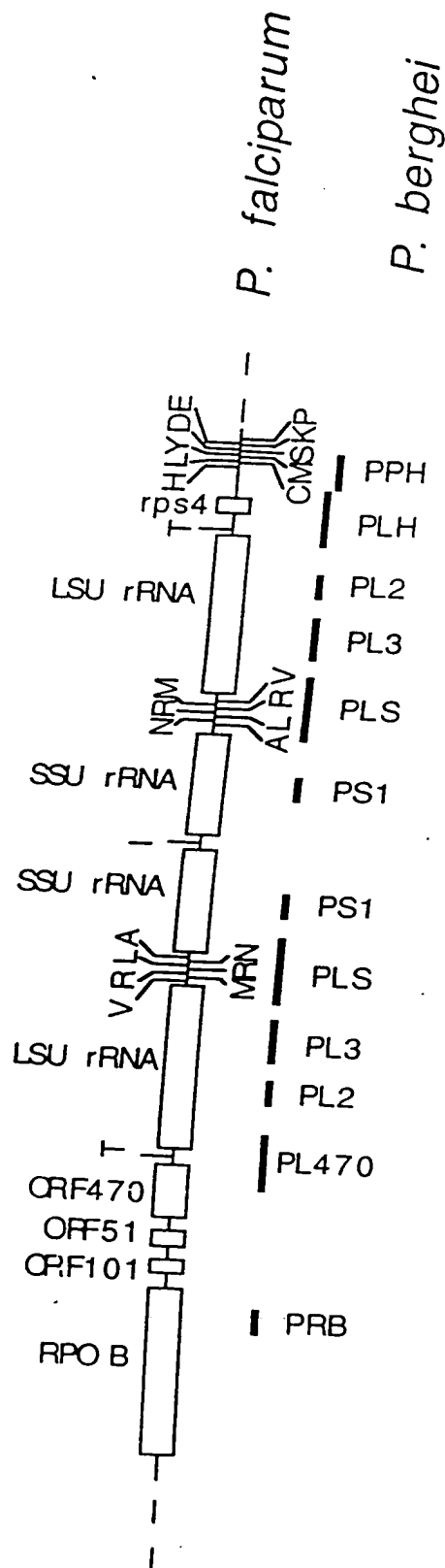


FIGURE 4B

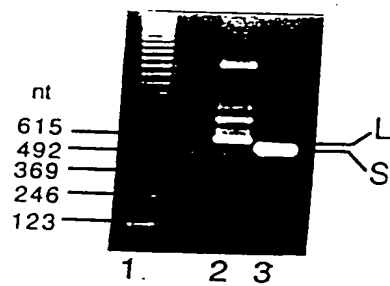


FIGURE 5